IN THE SPECIFICATION:

Please amend page 14, line 25 to page 15, line 8 as follows:

A DNA fragment containing the CRD region (ref.) of colicin E3 was amplified by PCR using primers represented by SEQ ID NO:1 and SEQ ID NO:2, and a DNA fragment containing the immunity (ref.) of the same using primers represented by SEQ ID NO:3 and SEQ ID NO:4, from an *E. coli* colicin E3 plasmid (pSH350) (which has been deposited on Jury 25, 2003, as FERM BP-8436 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan). Next, a DNA fragment represented by SEQ ID NO:7 was obtained by carrying out PCR, using a fragment prepared by fusing both of the fragments as the template and using primers represented by SEQ ID NO:5 and SEQ ID NO:6. The structure of this DNA fragment is shown below.

Please amend page 17, line 19 to page 18, line 25 as follows:

Each of these DNA fragments having 1 to 5 amber termination codons was subjected to TA cloning using pGEM T easy vector (manufactured by Promega), and then to sequence analysis to obtain 5 plasmids having respective insertion fragments of correct nucleotide sequences, namely pCI3A1 (which has been deposited on Jury 24, 2003, as FERM BP-8437 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and

Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan), pCI3A2 (which has been deposited on Jury 24, 2003, as FERM BP-8438 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan), pCI3A3 (which has been deposited on Jury 24, 2003, as FERM BP-8439 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan), pCI3A4 (which has been deposited on Jury 24, 2003, as FERM BP-8440 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan) and pCI3A5 (which has been deposited on Jury 24, 2003, as FERM BP-8441 in International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1, Higashi 1-Chome, Tsukuba-shi, Ibaraki-ken, Japan). On the other hand, as the vector, a plasmid pBS2SKP-SfiI into which two *Sfi*I cleavage sites (shown by underlines) having different protruding terminal sequences were inserted was constructed by annealing two synthetic single-stranded oligonucleotides represented by SEQ ID NO:21 and SEQ ID NO:22, and inserting the thus formed double-stranded DNA fragment between *Bam*HI and *Eco*RI of pBluescript II SK(+). This plasmid was digested with *Sfi*I, ligated with the above-described colicin E3 CRD gene fragments having 1 to 3 amber termination codons, and then transformed into an *E. coli* strain XL1-Blue by electroporation. As a result of spreading the thus obtained *E. coli* cell suspension on an agar medium containing 100 mg/l ampicillin and 0.1% glucose and culturing at 37°C for a whole day and night, transformants were obtained on the agar medium only in the case in which three amber termination codons were inserted. When plasmid pBS-Sfi-a3col was recovered from the thus obtained transformants and transformed into XL1-Blue, and then the resulting *E.*

coli cell suspension was spread on an agar medium containing 100 mg/l ampicillin + 0.1% glucose, and on an agar medium containing 100 mg/l ampicillin + 200 μM IPTG (isopropyl-β-D-thiogalactopyranoside) and cultured at 37°C for a whole day and night, a large number of colonies were formed only when cultured on the medium containing glucose, and formation of colonies was not found on the medium containing IPTG.